



Acrylamide in Food: Sources and Prevention

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Abstract

Acrylamide, a toxic compound with potential carcinogenic effects, is commonly formed in carbohydrate-rich foods through the Maillard reaction between reducing sugars and amino acids during high-temperature cooking processes. This study reviews the sources of acrylamide in food, its health risks, and various mitigation strategies. Acrylamide is prevalent in products such as bread, potato products, and coffee. Its formation is influenced by factors like pH, temperature, and cooking time. Effective mitigation strategies include modifying cooking methods, using enzymes like asparaginase, selecting low-carbohydrate materials, and applying pretreatments such as soaking in specific solutions or adding lactic acid bacteria. Regulatory measures by national governments and international organizations aim to minimize acrylamide levels in food to protect public health. Future research should focus on developing cost-effective and widely applicable methods to further reduce acrylamide content in food products.

Keywords: acrylamide, maillard reaction, health risk, prevention, mitigation strategies.

1 Introduction

Acrylamide (C_3H_5NO) is a prevalent chemical compound formed during the baking, frying, or roasting of carbohydrate-rich foods at temperatures exceeding 120°C , especially in low-moisture environments. This formation occurs through the Maillard reaction between reducing sugars (e.g., glucose, fructose) and amino acids (especially asparagine), contributing to non-enzymatic browning and imparting flavor and color to food [1–3]. Acrylamide is commonly found in bakery products such as bread, cereal, and potatoes, with the highest levels detected in roasted coffee, French fries, potato chips, and grains [4–6]. It is also used industrially as a thickener or flocculating agent in water treatment, cosmetics, and paper [7].

Despite its widespread presence and industrial applications, acrylamide poses significant health risks when present in food. It has been associated with several adverse health effects, including neurotoxicity, which can lead to nervous system damage, and reproductive toxicity, which can affect fertility and reproductive health. Additionally, acrylamide is classified as a probable human carcinogen by the International Agency for Research on Cancer (IARC), meaning it can increase the risk of developing cancer. There is also evidence suggesting that acrylamide may cause genotoxicity, which involves damage to the genetic information within a cell causing mutations [8]. The harmful effects of acrylamide were first identified in 2002 by researchers



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from the University of Stockholm and the Swedish National Food Administration[9]. The World Health Organization (WHO) regulates acrylamide levels in food, with daily exposure limits for adults ranging from 0.2-1.0 $\mu\text{g/kg}$ body weight [10].

In Europe, legislation such as recommendation 2007/331/EC monitors acrylamide levels in foods. The European Food Safety Authority (EFSA) adopted a statement in 2005, aligning with the Joint FAO/WHO Expert Committee on Food Additives (JECFA) risk assessment. A 'toolbox' guides producers on limiting acrylamide in food, with standards for nine food types across 27 EU countries. EFSA updates these recommendations based on market changes, such as the introduction of the 'other products' category in recommendation 2010/307/EC. In California, the Safe Water and Toxic Enforcement Act (1986) includes acrylamide among over 950 toxic substances, requiring companies to provide warnings if exposure levels exceed safe limits, defined as no more than one cancer case per 100,000 people over a lifetime of exposure.

In China, the Ministry of Health's 2005 Risk Assessment of Acrylamide in Food advises reducing fried and high-fat food consumption to mitigate health risks (www.gov.cn). Although specific laws are not yet in place, many large food companies have begun controlling acrylamide content, with regulations expected soon.

Acrylamide detection methods have evolved from derivatization and gas chromatography-mass spectrometry (GC-MS) [11, 12] to more advanced techniques like Liquid Chromatography-Tandem Mass Spectrometry (LC-MS/MS). Current methods include GC, High-Performance Liquid Chromatography (HPLC), and Mass Spectrometry, though these are often slow and costly. Alternatives like GC-ECD [12], HPLC with UV detection (BGVV, 2002), and reversed-phase HPLC-DAD [13] have been developed. Other methods include LC-UV [14–16], capillary electrophoresis (CE), solid-phase extraction (SPE), and dispersive liquid-liquid microextraction (DLLME) [17]. The European Committee for Standardization has published LC-ESI-MS/MS for determining acrylamide in bakery products.

Given the potential health risks, it is crucial to review and understand the sources, formation mechanisms, and mitigation strategies for acrylamide in food. This review aims to provide a comprehensive overview of current research on acrylamide, highlighting the importance of controlling its levels in food

products to protect public health. By examining various detection methods, regulatory measures, and mitigation strategies, this review seeks to inform future research and policy development aimed at reducing acrylamide exposure from dietary sources.

2 Formation of Acrylamide

According to prior research, there are two main pathways leading to the formation of acrylamide in food. The first is the Maillard reaction, a common reaction occurring during thermal treatment. The second is fat oxidation, where acrolein released by oxidative lipid degradation leads to acrylic acid formation [18].

2.1 Maillard reaction

The Maillard reaction, occurring between reducing sugars and amino acids during food processing, is crucial for flavor, browning, and aroma development. However, it can produce toxic by-products like acrylamide, a known carcinogen [7].

The primary pathway for acrylamide formation is the asparagine route. Asparagine and reducing sugars form Schiff bases, which decarboxylate to produce intermediates. The carboxylate ion in the Schiff base facilitates decarboxylation, leading to the removal of hydrogen from the hydroxy group and forming an alpha-hydroxy anion. This anion then removes the acidic alpha-hydrogen, resulting in acrylamide, as shown in Figure 1. The critical step is the decarboxylation of the Schiff base, leading to Maillard intermediates that release acrylamide [18].

Two main hypotheses explain this pathway. Mottram et al. [19] identified Strecker aldehyde as a key precursor, facilitated by alpha-dicarbonyls. Asparagine, abundant in potatoes and cereals, significantly influences acrylamide production, as shown in Figure 2. Stadler et al. [20] suggested that asparagine with alpha-dicarbonyls produces lower acrylamide levels compared to sugar/asparagine mixtures, but reactions with acetol yield the highest levels.

Stadler et al. [20] also proposed that early-stage Maillard reaction glycoconjugates, including N-glycosides, are pivotal in acrylamide formation. This hypothesis is supported by Yaylayan et al. [21] and Zyzak et al. [22]. Thus, while the asparagine route is primary, it involves various intermediates.

Alternative pathways exist. Figures 3 and 4 illustrate three:

1. Aspartic acid, carnosine, and beta-alanine enhance acrylamide formation through acrylic acid, which combines with ammonia to form acrylamide [20, 23].
2. Aminopropionamide, a dominant intermediate from asparagine, forms acrylamide through reactions with pyruvic acid [20, 22].
3. Pyruvic acid, produced by the dehydration and desulphurization of serine and cysteine, reduces to lactic acid and further dehydrates to acrylic acid, which then transforms into acrylamide [24, 25].

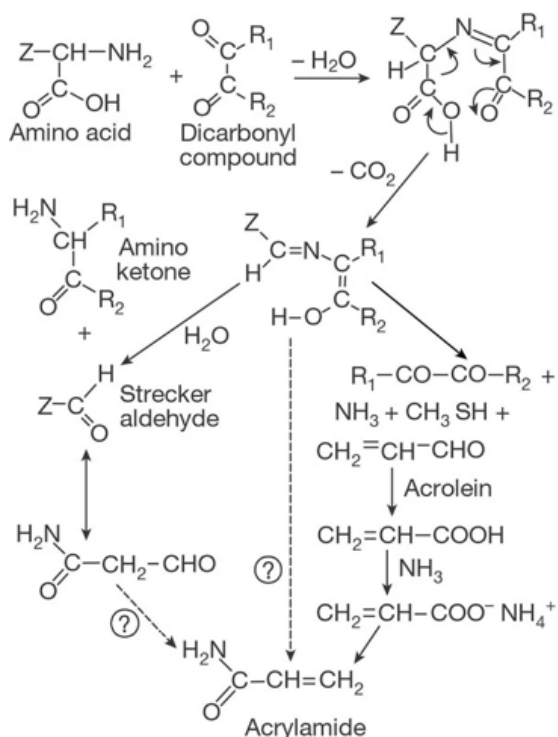


Figure 1. The reaction mechanism of the Maillard reaction between asparagine and reducing sugar, showing three intermediates during the reaction [27].

2.2 Fat oxidation

Besides the Maillard reaction, fat oxidation is another pathway for acrylamide formation, usually occurring in fatty acid-rich foods as shown in Figure 4. Fatty acids first change to acrolein through oxidative degradation, and then secrete acrylic acid to achieve acrylamide formation [20, 26]. Acrolein is a common aldehyde detected as a cytotoxic component [28]. In food, it is always present in oil and fat. There is also a new pathway showing that acrolein can react with asparagine to generate high levels of acrylamide under suitable conditions, indicating a critical role of acrolein in lipid-rich foods for acrylamide formation [29, 30]. Fat, together with nitrogen compounds in food, produces acrylamide after acrolein transfers to acrylic

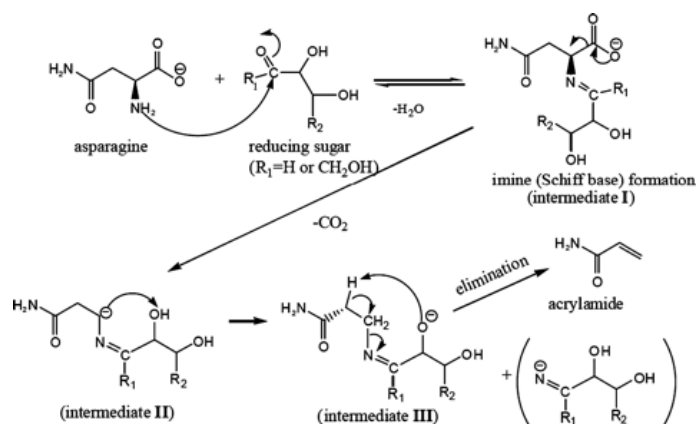


Figure 2. Possible pathway for acrylamide formation after Strecker degradation of the amino acids asparagine. In asparagine, the side chain Z is $-CH_2CH_2SNH_3$, while in methionine, Z means $-CH_2CH_2SNH_3$ [19].

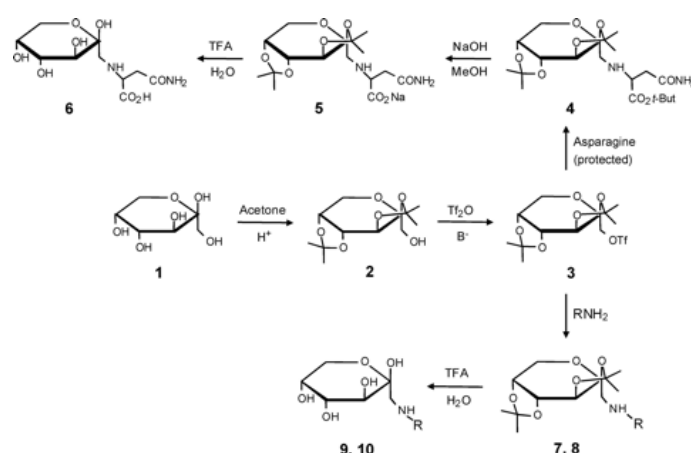


Figure 3. Synthetic pathway to the Amadori compounds of asparagine [20].

acid. Then, there is a reaction between acrylic acid and ammonia, which can be achieved by the pyrolysis of nitrogen-containing compounds.

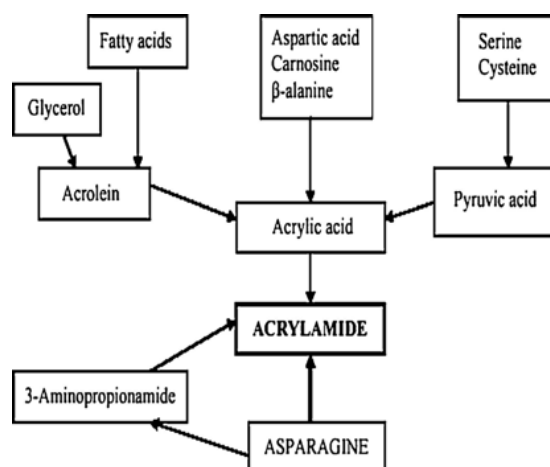


Figure 4. Alternative formation routes of acrylamide via the Maillard reaction system [27].

3 The Factors Affect the Formation of Acrylamide in Industrial Process

3.1 pH

In industrial processes, pH plays a crucial role in many processing steps and in inhibiting the formation of certain compounds. For acrylamide (AM) formation, it is also a pH-dependent compound. Different types of food have different formation mechanisms, resulting in varying pH ranges to inhibit acrylamide.

Charoenprasert et al. [31] illustrated that pH value can influence the acrylamide content in California-style black ripe olives (CBROs). Commercial olives contain high levels of acrylamide, so [31] tried to find a method to reduce these levels while preserving the olives' sensory texture. As mentioned in the formation section, acrylamide easily forms in potatoes and grains, which contain high levels of sugar. These food materials have high levels of acrylamide precursors. However, olives do not have these components, being a fatty acid-rich fruit. The exact formation mechanism in olives is still unknown, but studies suggest it is correlated with the processing step—sterilization [32]. Charoenprasert et al. [31] demonstrated that glucosamine levels before sterilization and acrylamide concentration after processing are significantly positively correlated. In their research, [31] added sodium hydroxide ($NaOH$, pH over 12, 0.25 N) to facilitate the base-catalyzed degradation of glucosamine. According to Table 1, the higher the pH value of the solution treating the olives, the lower the acrylamide content in the CBROs, especially at pH values above 12, with results of $3.91b \pm 0.40$ and $2.17c \pm 0.20$ at pH 12 and pH 13, respectively. Additionally, the longer the olives were soaked in sodium hydroxide, the lower the acrylamide content, provided the soaking time was within 5 days. The details are shown in Figure 5.

Table 1. Effect of pH values of $NaOH$ on the acrylamide concentration in California-style black ripe olives.

Treatment solutions	Acrylamide (nmol/g)*
Control (H_2O)	$8.24a \pm 0.28$
Solution of sodium hydroxide, pH 9	$7.79a \pm 0.29$
Solution of sodium hydroxide, pH 10	$8.31a \pm 0.62$
Solution of sodium hydroxide, pH 11	$7.93a \pm 0.26$
Solution of sodium hydroxide, pH 12	$3.91b \pm 0.40$
Solution of sodium hydroxide, pH 13	$2.17c \pm 0.20$

*Malues with different letters are significantly different at 95% confident level.

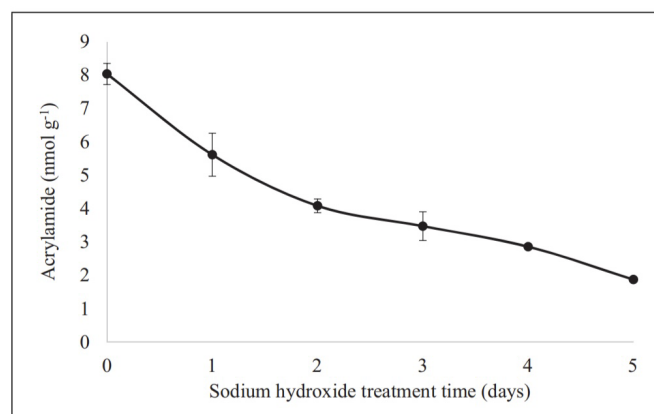


Figure 5. Effect of $NaOH$ treatment time on concentration acrylamide in California-style black ripe olives.

influencing acrylamide content in wholegrain biscuits and cocoa biscuits. Biscuits are a common snack made by heating dough, which contains high levels of reducing sugars such as glucose and sucrose. Based on the mechanism, it is easy to form acrylamide in biscuits due to their carbohydrate-rich materials and high levels of precursors. In this case, pH value, heating time, and temperature are relevant factors affecting acrylamide levels. When the pH increases, the reaction between reducing sugars and asparagine is enhanced. Therefore, Suman et al. [33] emphasized the importance of adding acidification agents to biscuit dough. In the food industry, citrate and tartaric acid are commonly used as acidification agents, while biscuits need to be fluffy. This is why the authors added raising agents like sodium bicarbonate or ammonium hydrogen carbonate. Both raising agents and acidification agents help decrease acrylamide levels. Commercially, biscuits should taste good. Ensuring an acceptable range from a sensory perspective, there is a drop in acrylamide levels by more than 50% as the pH value and sodium bicarbonate concentration change. Suman et al. [33] illustrated that $NaHCO_3$ is more effective in limiting acrylamide formation compared to other raising agents. Graf et al. [34] determined that acrylamide levels reduce by almost 70% when sodium bicarbonate is applied.

Yuan et al. [35] showed the effects of pH value on acrylamide formation in potato chips. Although this two thermal methods (conventional heating and microwave) and the type of sugar (fructose and glucose) reacting with asparagine, their research also demonstrated higher levels of acrylamide when the pH value is high, treated by these two methods. The graph shows no acrylamide content detected when

In the biscuit industry, pH also plays a significant role. Suman et al. [33] conducted research on factors

pH is equal to 3. This may relate to the form of asparagine and reducing sugars, where they present in a protonated form [36]. During the pH range from 4 to 8, acrylamide content increases in the chips. The formation rate reaches its highest when the pH value climbs to 8 in both fructose and glucose systems. This matches previous studies by Brown et al. [37] and Rydberg et al. [38], which report the maximum formation rate at pH 7 to 8. Surprisingly, acrylamide content falls after the pH rises to 10. This trend is similar to acrylamide in biscuits, as both contain high concentrations of reducing sugars.

3.2 Time for Heating

The time of thermal treatment is critical in industrial processes, including acrylamide formation. Heating time influences acrylamide levels until a maximum point is reached [18]. The type of food also affects the required heating time.

Suman et al. [33] found that baking time significantly affects acrylamide levels in biscuits, with longer times promoting its formation, though pH is more critical than time and temperature. Deoxynivalenol (DON), a common contaminant in wholegrain bakery products, is also influenced by baking time. The greatest effect was observed when baking biscuits at 200°C for 8 minutes. Increasing baking time effectively reduces DON contamination [39]. Reducing baking time to 5 minutes resulted in a 77-100% reduction in acrylamide levels while maintaining good sensory qualities. Thus, shorter baking times result in lower acrylamide levels in biscuits. To control both DON and acrylamide, the time-temperature parameters of baking are crucial. Baking at 180°C for 5 minutes is optimal for cocoa and wholegrain biscuits, achieving acrylamide contents of $43 \pm 13 \mu\text{g/kg d.m.}$ and $16 \pm 3 \mu\text{g/kg d.m.}$, respectively, while minimizing DON content.

For potato chips, Tareke et al. [40] and Becalski et al. [41] demonstrated that longer frying times increase acrylamide levels. Yuan et al. [35] found that acrylamide content generally increased with longer treatment times, regardless of pH and heating methods. Acrylamide levels rose significantly after 20 minutes at pH 8.0, reaching $3.0 \mu\text{g/ml}$ at 30 minutes [42]. Esposito et al. [43] recommend that control the frying time in 0-4mins to minimize acrylamide formation as shown in Figure 6.

In roasted coffee, acrylamide levels are high due to precursors in coffee beans. In Figure 7, Santos et al. [44] reported that roasting at 200°C for less than

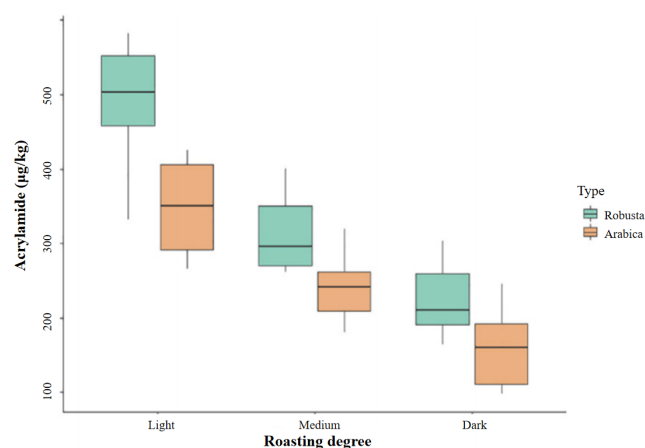


Figure 6. Change in acrylamide content of potato chip during frying at 170 degrees.

20 minutes is optimal. Acrylamide levels peak early in the roasting process and then decrease due to degradation or depletion of precursors. Light coffee, especially Robusta, has the highest acrylamide levels ($500 \mu\text{g/kg}$), while Arabica has the lowest ($150 \mu\text{g/kg}$) [4, 81].

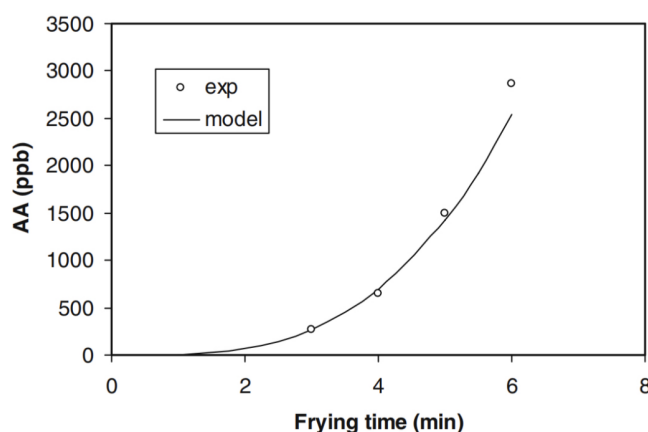


Figure 7. Distribution of occurrence of acrylamide in Arabica and Robusta coffee.

Time also affects acrylamide levels in starch-based and cereal systems, such as bread and flatbread. Acrylamide levels peak at approximately 200°C, depending on the system and baking time [48]. Longer baking times reduce acrylamide levels, similar to dark coffee. For example, flatbread baked at 260°C for 17.5 minutes had the lowest acrylamide levels under 20 mg/kg. Different conditions for bread crusts achieved similar acrylamide content, such as 195°C for 19.4 or 40.6 minutes, and 230°C for 15 minutes.

3.3 Temperature applied

Temperature is crucial in processing, significantly influencing acrylamide levels in food. Higher

temperatures in starch-based systems result in higher acrylamide content [48]. Heating temperature combined with time is critical.

In the biscuit industry, Suman et al. [33] found that baking at 200°C increased acrylamide in wholegrain and cocoa biscuits. Reducing the temperature to 180°C dropped acrylamide levels to below $125 \pm 14 \mu\text{g/kg}$ d.m. and $156 \pm 15 \mu\text{g/kg}$ d.m., compared to $306 \pm 16 \mu\text{g/kg}$ d.m. and $400 \pm 27 \mu\text{g/kg}$ d.m. at 200°C. The optimal temperature for baking biscuits is 180°C with a 5-minute baking time, achieving a 77-100% reduction while maintaining sensory quality.

For coffee roasting, Santos et al. [44] reported that roasting is usually done at around 200°C. Optimal final temperatures for different roasting levels and coffee types vary between 215-238°C [43]. Acrylamide forms during frying, roasting, and baking above 120°C but degrades at temperatures as low as 180°C. The effect of temperature on acrylamide levels is closely related to heating time [45, 46].

For potato chips, high temperatures lead to acrylamide formation, with reducing sugars as limiting precursors [50, 51]. Gokmen et al. [52] found acrylamide levels rise substantially with frying temperature, reaching almost 6000 ppb at 190°C compared to 1000 ppb at 150°C. Knol et al. [47] showed that glucose converts to fructose above 160°C, and the reaction between fructose and asparagine is preferred over glucose above 140°C. High temperature and carbohydrate conversion significantly influence acrylamide formation. During frying, oil temperature above 170°C allows temperature increase and water evaporation, favoring acrylamide formation [52].

Temperature also affects acrylamide formation in starch-based and cereal systems. Tareke et al. [40] showed acrylamide levels increase with baking temperature. Bräthen et al. [49] indicated that baked starch gel changes color from white to dark as temperature increases. Acrylamide content in freeze-dried flatbread peaks at 180°C with a 5-minute baking time, reaching almost 3600 mg/kg. Ordinary flatbread baked at 200-220°C achieves the highest acrylamide content. At temperatures above 260°C, acrylamide concentration peaks at nearly 500 mg/kg in bread crusts. Acrylamide reduction in starch model systems is controlled by time-temperature combinations.

4 The Mitigation Strategies for Preventing Acrylamide

To reduce acrylamide levels in food, researchers have explored various methods. These methods are compiled into a 'toolbox' to assist food companies and other researchers in acrylamide reduction efforts. This contributes to global food safety by helping residents understand the toxicity and hazards of acrylamide.

4.1 Different Cooking Methods

Different cooking methods are crucial in mitigating acrylamide in food, as they can produce food products that are sensory acceptable to consumers. This section introduces conventional heating methods, microwave cooking, and air-frying. Conventional heating methods, such as boiling, frying, steaming, and baking, have been used for many years. However, they do not significantly reduce acrylamide levels in food. For instance, the acrylamide content in roasted coffee powder ranges from $135 \mu\text{g/kg}$ to $1139 \mu\text{g/kg}$, while French fries contain $36 \mu\text{g/kg}$ to $1441 \mu\text{g/kg}$, and potato chips range from $211 \mu\text{g/kg}$ to $3515 \mu\text{g/kg}$ [53]. Boiling provides the lowest acrylamide levels in food. At a pH of 8.0, the optimal condition for acrylamide formation, the acrylamide content in potato chips ranges from $0.5 \mu\text{g/ml}$ to $0.9 \mu\text{g/ml}$, which is lower than when using microwaves.

4.1.1 Microwave

Microwave cooking, developed by Raytheon engineer Percy Spencer in 1945, is a widely used thermal method. Yuan et al. [35] found that microwave ovens can also be used in potato chip production. Under the same pH conditions, lower microwave power results in less acrylamide formation. This reduction can be monitored by controlling the microwave power. Compared to traditional frying (180°C for 4 minutes), microwaving at 650W for 3 minutes reduces acrylamide levels by $100 \mu\text{g/kg}$. Sansano et al. [54] reported similar findings, with acrylamide levels decreasing when microwaving at 600W for 1 to 5 minutes. For instance, the acrylamide level is $63(5) \mu\text{g/kg}$ using deep-oil frying for 2 minutes, but decreases to $25(5) \mu\text{g/kg}$ with microwave frying. The formation of acrylamide by microwave frying appears to depend on the frying technique. Except at the end of frying, the acrylamide content of potatoes is higher than that of microwave-fried potatoes. During the microwave process, acrylamide levels increase exponentially in potato products [54]. However, some studies found that microwaving might favor acrylamide formation in some frozen par-fried

French fries [55]. Therefore, specific conditions are necessary to effectively reduce acrylamide content using microwaves.

4.1.2 Air-frying

Air-frying is a recent cooking technique that has gained popularity in households. It is a small countertop convection oven designed to simulate deep frying without adding extra oil. It can be used to bake cakes, make French fries or potato chips, and roast chicken wings. Sansano et al. [56] conducted a study comparing air-frying and deep-oil frying at 180°C. The results show that, although the Maillard reaction rate is faster for deep-frying, air-frying reduces acrylamide levels by 90% without any pretreatment in fried potatoes. This is due to differences in heat transfer between the two thermal technologies. With both mediums, oil and air, at 180°C, oil is more likely to conduct heat to the potato surface [57]. In [54]'s studies, the CIE Lab* parameters were used to present acrylamide content, as the color of the potato changes with increasing acrylamide content. Frying technology most affected the variation of color parameters during cooking, especially a*. The L* value of air-frying remains almost constant, while deep-oil frying results in darker potatoes. The b* value indicates that deep-oil fried potatoes are more yellow than air-fried ones. Thus, air-frying can be used instead of traditional oil frying to reduce acrylamide levels in food products, specifically potato chips or French fries.

4.1.3 Vacuum frying

Vacuum frying is a new technique involving deep-frying and dehydrating food at low temperatures (80-120°C), which effectively reduces nutrient damage caused by high temperatures. Garayo et al. [58] reported that vacuum frying could be an alternative method to produce potato chips with lower acrylamide and oil content, and better texture. The study used temperatures of 118°C, 125°C, and 140°C at a vacuum pressure of 10 Torr. Acrylamide levels decreased by 63% from 140°C to 125°C and by 10% from 125°C to 118°C. Compared to traditional frying, vacuum frying at 118°C reduces acrylamide content in potato slices, resulting in golden color and satisfactory texture. The highest acrylamide content by vacuum frying is over 500 ppb, while traditional frying results in almost 800 ppb. The CIELab* parameters show that vacuum-fried potato chips have lower acrylamide levels than traditionally fried chips [59–61]. Overall, vacuum frying can replace traditional oil frying for making potato chips.

4.1.4 Best cooking method

Among the cooking methods for preventing acrylamide formation, air-frying is the most effective. Studies have shown that air-frying can reduce acrylamide levels in fried potatoes by up to 90% without any pretreatment [56]. Compared to traditional oil frying, air-frying uses hot air to cook food, which reduces the likelihood of high temperatures that promote acrylamide formation while maintaining food quality and sensory properties [57]. Additionally, air-frying reduces the overall fat content in food, making it a healthier cooking method. Therefore, air-frying is recommended as the best cooking method for reducing acrylamide formation.

4.2 Enzyme

Asparaginase is an enzyme present in living organisms, plants, and animals [62]. It hydrolyzes asparagine to aspartic acid and ammonia [63], reducing acrylamide formation by decreasing asparagine levels. The enzyme is derived from sources like *E. coli*, *A. oryzae*, and *A. niger*. Researchers have used asparaginase to reduce acrylamide in French fries, bread, potato chips, and biscuits. Zyzak et al. [22] used *E. coli* asparaginase, achieving a 99% reduction in acrylamide in potatoes. Ciesarová et al. [64] treated potatoes at 180°C for 20 minutes, resulting in a 50-90% reduction. Mahajan et al. [65] used *Bacillus licheniformis*, achieving an 80% reduction. For French fries, Pedreschi et al. [66] used *A. oryzae*, achieving a 67% reduction at 175°C for 3 minutes. Zuo et al. [67] used *Thermococcus zilligii*, achieving an 80% reduction at 175°C for 5 minutes. For potato chips, Hendriksen et al. [63] showed that 10,500 ASNU/L of asparaginase reduced acrylamide from 1700 to 700 µg/kg (60% reduction). Pedreschi et al. [68] achieved a 90% reduction at 170°C for 5 minutes. Onishi et al. [69] used *Bacillus subtilis*, achieving an 80% reduction. For bread, Tuncel et al. [70] used *Pisum sativum* L., achieving a 57-68% reduction at 220°C for 22-25 minutes. Ciesarová et al. [71] used *A. niger*, achieving up to a 90% reduction. Kumar et al. [72] used *Cladosporium* sp., achieving a 97% reduction in crust and 73% in crumbs at 220°C for 25 minutes. Amrein et al. [73] used *E. coli*, achieving a 55% reduction in gingerbread. Huang et al. [74] used *Rhizomucor miehei*, achieving a 94.2% reduction in biscuits at 200°C for 25 minutes. Anese et al. [75] used different enzyme dosages in biscuits, achieving a 69% reduction at 200°C. Hendriksen et al. [63] used *A. oryzae*, achieving a 65-84% reduction in semi-sweet biscuits and 34-90% in ginger biscuits. The optimal

conditions were pH 7.0 and 40°C, with 1050 ASNU/kg of enzyme reducing acrylamide to 10 µg/kg compared to 450 µg/kg in controls.

4.3 Reducing Sugar

As mentioned, acrylamide formation is strongly related to reducing sugars. Researchers have indicated that reducing sugar concentration plays a key role in acrylamide formation. Therefore, decreasing reducing sugar levels should lower acrylamide content. The type of reducing sugar (glucose, fructose, sucrose) also affects acrylamide levels in carbohydrate-rich foods.

In industrial biscuit production, Suman et al. [33] reported that glucose significantly increases acrylamide levels. Using high dextrose levels at 200°C for 8 minutes increased acrylamide by 120% compared to controls. Conversely, a low glucose concentration combined with suitable time-temperature conditions (180°C, 8 minutes) resulted in a 77% decrease in acrylamide. Using sucrose as the reducing sugar source in biscuits can decrease acrylamide production by about 50% [34, 76]. Graf et al. [34] noted that low acrylamide levels form at 160°C, remaining below 150 µg/kg, and recommended using sucrose at 180°C for biscuit production.

In the potato products industry, Yuan et al. [35] found that acrylamide levels were lower when using glucose compared to asparagine-fructose combinations. At pH 8.0, the optimal condition for acrylamide formation, the acrylamide level was 1.0 µg/ml for Asn & Fru boiled at 120°C for 30 minutes, while it was 0.52 µg/ml for Asn & Glc combinations. At pH 3.0, acrylamide generation was almost zero, regardless of the combination. Muttucumaru et al. [77] showed that the ratio of asparagine to reducing sugar is crucial for acrylamide formation in potato products, with a tipping point value of 2.257 ± 0.149 . Potatoes in Doncaster had higher free asparagine levels than in Woburn, resulting in lower average acrylamide levels among 20 types. Reducing sugar levels increased with storage time, but acrylamide levels were not highest in Harmony potatoes due to their lower asparagine-to-reducing sugar ratio.

Liyanage et al. [78] found that the percent decrease in asparagine content (1–60%) was generally lower than that in reducing sugars (28–94%) with increasing frying time at a constant temperature. Amrein et al. [50] reported that acrylamide formation is primarily limited by available reducing sugars rather than free asparagine. Elmore et al. [79] found that only 0.29%

of asparagine was converted to acrylamide at 180°C, while reducing sugars were dominant in acrylamide formation. Bread contains various reducing sugars, including glucose, fructose, and maltose, which contribute to acrylamide formation. Crawford et al. [80] identified sugar as a factor in acrylamide formation in commercial flatbread but lacked accurate data for analysis.

In coffee products, different coffee beans contain varying percentages of asparagine and reducing sugars, resulting in different acrylamide levels. Arabica coffee has lower acrylamide levels than Robusta due to its lower asparagine content at similar roasting degrees [43, 81].

4.4 Pretreatment before Processing

Pretreatment is a crucial step before processing, involving a series of cleaning operations to remove impurities that adversely affect dyeing and printing. For example, soaking materials in a solution or adding a solution to the materials can be beneficial. Pre-thawing frozen materials to change their form is also a type of pretreatment. Various pretreatment methods are used to reduce acrylamide levels, depending on the type of food. For instance, significant research has focused on potato products.

4.4.1 Microwave Pre-thawing

Microwave pre-thawing is a method used to thaw frozen food. Due to commercial demands, many potato strips are frozen. Some authors suggest that pre-thawing frozen potato strips to decrease frying time may be an effective approach to reduce acrylamide levels [82]. The authors fried potato slices at 170°C, 180°C, and 190°C after pre-thawing. The acrylamide level dropped by 10% after pre-thawing the potato slices, from 17.7 to 15.9 ng/g. The researchers recommend frying at 180°C after pretreatment, as this can reduce acrylamide levels by 89%, a significant decrease, from 72.1 to 8 ng/g. When frying at 190°C, the reduction in acrylamide was 64%, from 50.5 to 18.4 ng/g. The combination of pre-thawing and heating temperature is crucial for acrylamide reduction. When frying at 170°C, the acrylamide level showed a slight decrease, possibly because acrylamide formation is not accelerated at this temperature. Previous studies indicated that the acrylamide formation rate increases significantly above 175°C [26]. This study also examined the quality of the final products after treatment and frying. Lab* data showed that all control samples were slightly more yellow than the pre-thawed ones, which may be more appealing to

consumers. All data were compared with control samples fried at the same temperature without pre-thawing.

4.4.2 Hydrocolloids

Hydrocolloids are polysaccharides connected by glycosidic bonds, consisting of more than ten monosaccharides. Polysaccharides can be classified as homo- and hetero-polysaccharides. Homo-polysaccharides consist of identical monosaccharides, such as cellulose and amylose. In contrast, hetero-polysaccharides consist of different monosaccharides (at least two types), including alginate and xanthan. Hydrocolloids have a high affinity for aqueous solutions and are used in various industries. They can function as thickeners, gelling agents, foam stabilizers, emulsifiers, and dispersants, inhibit the formation of ice and sugar crystals, and control the release of flavors.

In 2020, Liu et al. [83] demonstrated that using sodium alginate as a coating for fried potato chips can reduce acrylamide levels in food products. Alginate is a polysaccharide extracted from seaweed. After acidification, alginic acid, which is soluble in $NaOH$ or Na_2CO_3 , is converted to sodium alginate, an anionic polymer. Liu et al. [83] determined that a sodium alginate concentration of 1.34%, frying at 179°C for 4.38 minutes, was optimal. The evidence showed that sodium alginate significantly reduced acrylamide levels, inhibiting up to 76.59% compared to control samples. The sodium alginate coating effectively prevented potato chips from absorbing oil while maintaining good quality and sensory properties. Due to the poor solubility of sodium alginate, researchers used low dosages ranging from 0.2% to 2.2%. They found that around 1.4% sodium alginate provided the best inhibition rate, reaching 58.93%. Sodium alginate contains M-blocks and G-blocks, both contributing to the viscosity and thickening of products, which explains the reduced oil absorption. Sensory evaluations indicated that coated potato chips tasted similar to regular ones. Sodium alginate also enhances satiety signals to the brain, reducing blood cholesterol, glucose levels, and total daily calorie intake. Data from several studies suggest that other hydrocolloids, such as guar gum and pectin, also reduce oil uptake during frying [84, 85]. However, sodium alginate is the most effective coating.

Champrasert et al. [86] reported that alginate, pectin, and chitosan positively affect acrylamide reduction. Pectin is a complex group of plant

polysaccharides, consisting of galacturonic acid, rhamnose, arabinose, xylose, and other sugars, commonly added to fruit juice or jelly depending on the esterification of galacturonate residues. Chitosan, a linear polysaccharide, consists of glucosamine and GlcNAc randomly distributed and connected through β -glycosidic bonds. In this study, polysaccharides were used as pretreatment solutions for soaking potatoes. The potato products were fried at 180°C or microwaved at 800W for 60 seconds. Under conventional heating, the inhibition rate of acrylamide was effective, at 65% for alginate and 56% for pectin. However, the reduction decreased under microwave heating, to 36% and 30%, respectively, as high-efficiency microwaves may promote acrylamide formation. Additionally, Champrasert et al. [86] coated potatoes with alginate, pectin, and chitosan at the same concentration, frying at 170°C for 3 minutes and microwaving at 800W. The reduction of acrylamide was 53.5%, 51.2%, and 40.9%, respectively, when frying. When microwaved for 60 seconds at 800W, the reduction was slight, only 5%, with no reduction for chitosan-coated potatoes. Microwave irradiation may destroy the coating, preventing it from avoiding the temperature increase that forms acrylamide.

4.4.3 Other Soaking Solution

Sansano et al. [56] demonstrated that various chemical agents can be used as pretreatment solutions to reduce acrylamide levels, such as citrate, glycine, calcium lactate, $NaCl$, or nicotinic acid (Vitamin B3). The authors controlled the conditions under both air-frying and deep-oil-frying at 180°C. For oil-frying, the most effective method was soaking in 2% citric acid, which reduced acrylamide levels by almost 91% (from 125 $\mu\text{g}/\text{kg}$ to 11 $\mu\text{g}/\text{kg}$) compared to the control sample. For air-frying, 2% calcium lactate was the most effective pretreatment solution, providing a 75% decrease in acrylamide levels (from 29 $\mu\text{g}/\text{kg}$ to 7 $\mu\text{g}/\text{kg}$). The reduction is attributed to the decrease in reducing sugars after pretreatment. Interestingly, some pretreatments did not help reduce acrylamide levels. For example, the acrylamide level in samples treated with 1% $NaCl$ was 50% higher in air-frying compared to the control sample. This phenomenon is due to an increase in reducing sugar content after treatment with 1% $NaCl$, from 0.20(0.03)d to 0.21(0.02)d. Similar results were observed in samples pretreated with 1% sodium chloride solution, blanching, soaking in 1% or 2% citric acid, 2% $NaCl$, and 1% calcium lactate when using air-frying. $NaCl$, commonly known as salt, is an ionic compound, and

calcium lactate is a salt consisting of two lactate anions with each Ca cation. Another reason for the increase in acrylamide levels is that the presence of these mono- or divalent cations (Ca^{2+} or Na^{+}) on the product surface could reduce the water activity of the surface, favoring acrylamide formation in the fried product. However, in traditional oil-frying, as long as the potato strips were pretreated, the chips had lower acrylamide levels than the control samples.

4.5 Lactic Acid Bacteria

Lactic acid bacteria (LAB) are Gram-positive, low GC, acid-tolerant, mostly non-sporulating, non-respiratory bacilli or cocci. Found in decomposing plants and dairy products, LAB primarily produce lactic acid from sugar fermentation, historically used to increase acidity and prevent spoilage in food fermentations. Bread, a carbohydrate-rich food, requires fermentation, and several studies have investigated LAB starters to reduce acrylamide levels.

Bartkiene et al. [87] demonstrated that different LAB strains (*P. acidilactici*, *L. plantarum*, *L. curvatus*, and *L.(+)* lactic acid bacteria) grown at 30°C and 37°C showed varying acrylamide reductions. Dastmalchi et al. [88], and acrylamide formation in bread is influenced by many factors, especially reducing sugars [89]. However, *L. plantarum* significantly reduced acrylamide concentrations in bread samples. Whole rye flour, rich in cell walls, undergoes hydrolysis and solubilization during sourdough fermentation, facilitated by LAB enzymes. *L. plantarum* exhibited the lowest pH and highest amylolytic enzyme activity, likely contributing to the greatest acrylamide reduction (12 µg/kg dw) compared to the control (15 µg/kg dw).

Bartkiene et al. [87] also reported that LAB combinations can reduce acrylamide in bread. The most effective combination, *L. plantarum* and *P. pentosaceus*, reduced acrylamide levels to 4.01 ± 0.31 µg/kg, with the lowest level at 3.70 µg/kg in wheat bread. Additionally, this combination had the lowest a^* value among all samples.

5 Conclusion

This article summarizes the sources of acrylamide formation and preventive measures. Acrylamide is a toxic compound that may have carcinogenic effects on the human body. It is mainly produced through the Maillard reaction between sugars and amino acids. To protect public health, governments and international organizations have enacted relevant regulations to limit the content of acrylamide in food.

The article points out that future research should focus on finding economical and widely applicable methods to reduce acrylamide content, while controlling key production steps. For example, exploring the effects of using colloids in foods that are prone to acrylamide formation (such as coffee and bread), or studying whether pre-treatment of bread can reduce the acrylamide content after baking. In addition, extracting acrylamide from food for packaging or other industrial uses may also be a future research direction, although there are still certain challenges.

In summary, by controlling key factors such as temperature, time, and pH, as well as adopting strategies like enzyme treatment, alternative cooking methods, and reducing carbohydrate content, the acrylamide content in food can be effectively reduced to protect consumer health. Future research should further explore more economical and efficient methods to reduce acrylamide.

Data Availability Statement

Data will be made available on request.

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Conflicts of Interest

The author declare no conflicts of interest.

Ethical Approval and Consent to Participate

Not applicable.

References

- [1] Mesías, M., Morales, F. J., & Delgado-Andrade, C. (2019). Acrylamide in biscuits commercialised in Spain: A view of the Spanish market from 2007 to 2019. *Food & function*, 10(10), 6624-6632. [CrossRef]
- [2] Andačić, I. M., Tot, A., Ivešić, M., Krivohlavek, A., Thirumdas, R., Barba, F. J., ... & Brnčić, S. R. (2020). Exposure of the Croatian adult population to acrylamide through bread and bakery products. *Food chemistry*, 322, 126771. [CrossRef]
- [3] Mousavi Khaneghah, A., Fakhri, Y., Nematollahi, A., Seilani, F., & Vasseghian, Y. (2022). The concentration of acrylamide in different food products: a global systematic review, meta-analysis, and meta-regression. *Food Reviews International*, 38(6), 1286-1304. [CrossRef]
- [4] Mesías, M., & Morales, F. J. (2015). Acrylamide in commercial potato crisps from Spanish market: Trends from 2004 to 2014 and assessment of the dietary

- exposure. *Food and Chemical Toxicology*, 81, 104-110. [CrossRef]
- [5] EFSA Panel on Contaminants in the Food Chain (CONTAM). (2015). Scientific opinion on acrylamide in food. *Efsa Journal*, 13(6), 4104. [CrossRef]
- [6] Esposito, F., Nardone, A., Fasano, E., Triassi, M., & Cirillo, T. (2017). Determination of acrylamide levels in potato crisps and other snacks and exposure risk assessment through a Margin of Exposure approach. *Food and Chemical Toxicology*, 108, 249-256. [CrossRef]
- [7] Sarion, C., Codina, G. G., & Dabija, A. (2021). Acrylamide in Bakery Products: A Review on Health Risks, Legal Regulations and Strategies to Reduce Its Formation. *International Journal of Environmental Research and Public Health*, 18(8), 4332. [CrossRef]
- [8] Cantrell, M. S., & McDougal, O. M. (2021). Biomedical rationale for acrylamide regulation and methods of detection. *Comprehensive Reviews in Food Science and Food Safety*, 20(2), 2176-2205. [CrossRef]
- [9] Rifai, L., & Saleh, F. A. (2020). A review on acrylamide in food: Occurrence, toxicity, and mitigation strategies. *International journal of toxicology*, 39(2), 93-102. [CrossRef]
- [10] JECFA, W. (2011). Evaluation of certain food additives and contaminants: seventy-fourth report of the joint FAO/WHO expert committee on food additives: world health Organization. [CrossRef]
- [11] Andrawes, F., Greenhouse, S., & Draney, D. (1987). Chemistry of acrylamide bromination for trace analysis by gas chromatography and gas chromatography—mass spectrometry. *Journal of Chromatography A*, 399, 269-275. [CrossRef]
- [12] United States. Congress. House. Committee on Commerce. Subcommittee on Health and the Environment. (1996). Food Quality Protection Act of 1995: Hearings before the Subcommittee on Health and Environment of the Committee on Commerce, House of Representatives, One Hundred Fourth Congress, First Session, on H.R. 1627, June 7 and 29, 1995 (Vol. 4). U.S. Government Printing Office.
- [13] Gökmen, V., & Şenyuva, H. Z. (2006). A simplified approach for the kinetic characterization of acrylamide formation in fructose-asparagine model system. *Food Additives and Contaminants*, 23(4), 348-354. [CrossRef]
- [14] Arámbula-Villa, G., Flores-Casamayor, V., Velés-Medina, J. J., & Salazar, R. (2018). Mitigating effect of calcium and magnesium on acrylamide formation in tortilla chips. *Cereal Chemistry*, 95(1), 94-97. [CrossRef]
- [15] Lindsay, R. C., & Jang, S. (2005). Model systems for evaluating factors affecting acrylamide formation in deep fried foods. In *Chemistry and safety of acrylamide in food* (pp. 329-341). Boston, MA: Springer US. [CrossRef]
- [16] Saraji, M., & Javadian, S. (2019). Single-drop microextraction combined with gas chromatography-electron capture detection for the determination of acrylamide in food samples. *Food Chemistry*, 274, 55-60. [CrossRef]
- [17] Norouzi, E., Kamankesh, M., Mohammadi, A., & Attaran, A.M. (2018). Acrylamide in bread samples: Determining using ultrasonic-assisted extraction and microextraction method followed by gas chromatography-mass spectrometry. *Journal of Cereal Science*, 79, 1-5. [CrossRef]
- [18] Keramat, J., LeBail, A., Prost, C., & Soltanizadeh, N. (2011). Acrylamide in foods: chemistry and analysis. A review. *Food and bioprocess technology*, 4, 340-363. [CrossRef]
- [19] Mottram, D. S., Wedzicha, B. L., & Dodson, A. T. (2002). Acrylamide is formed in the Maillard reaction. *Nature*, 419(6906), 448-449. [CrossRef]
- [20] Stadler, R. H., Robert, F., Riediker, S., Varga, N., Davidek, T., Devaud, S., ... & Blank, I. (2004). In-depth mechanistic study on the formation of acrylamide and other vinylogous compounds by the Maillard reaction. *Journal of agricultural and food chemistry*, 52(17), 5550-5558. [CrossRef]
- [21] Yaylayan, V. A., Wnorowski, A., & Perez Locas, C. (2003). Why asparagine needs carbohydrates to generate acrylamide. *Journal of agricultural and food chemistry*, 51(6), 1753-1757. [CrossRef]
- [22] Zyzak, D. V., Sanders, R. A., Stojanovic, M., Tallmadge, D. H., Eberhart, B. L., Ewald, D. K., ... & Villagran, M. D. (2003). Acrylamide formation mechanism in heated foods. *Journal of agricultural and food chemistry*, 51(16), 4782-4787. [CrossRef]
- [23] Yaylayan, V. A., Locas, C. P., Wnorowski, A., & O'Brien, J. (2004). The role of creatine in the generation of N-methylacrylamide: a new toxicant in cooked meat. *Journal of Agricultural and Food Chemistry*, 52(17), 5559-5565. [CrossRef]
- [24] Wnorowski, A., & Yaylayan, V. A. (2003). Monitoring carbonyl-amine reaction between pyruvic acid and α -amino alcohols by FTIR spectroscopy a possible route to Amadori products. *Journal of Agricultural and Food Chemistry*, 51(22), 6537-6543. [CrossRef]
- [25] Yaylayan, V. A., Perez Locas, C., Wnorowski, A., & O'Brien, J. (2005). Mechanistic pathways of formation of acrylamide from different amino acids. In *Chemistry and safety of acrylamide in food* (pp. 191-203). Springer US. [CrossRef]
- [26] Gertz, C., & Klostermann, S. (2002). Analysis of acrylamide and mechanisms of its formation in deep-fried products. *European Journal of Lipid Science and Technology*, 104(11), 762-771. [CrossRef]
- [27] Eriksson, S. (2005). *Acrylamide in food products: Identification, formation and analytical methodology* (Doctoral dissertation, Institutionen för miljökem). [CrossRef]
- [28] Casella, I. G., & Contursi, M. (2004). Quantitative Analysis of Acrolein in Heated Vegetable Oils by Liquid Chromatography with Pulsed Electrochemical

- Detection. *Journal of Agricultural and Food Chemistry*, 52(19), 5816–5821. [CrossRef]
- [29] Yasuhara, A., Tanaka, Y., Hengel, M., & Shibamoto, T. (2003). Gas Chromatographic Investigation of Acrylamide Formation in Browning Model Systems. *Journal of Agricultural and Food Chemistry*, 51(14), 3999–4003. [CrossRef]
- [30] Zhang, Y., Zhang, G., & Zhang, Y. (2005). Occurrence and analytical methods of acrylamide in heat-treated foods. *Journal of Chromatography A*, 1075(1–2), 1–21. [CrossRef]
- [31] Charoenprasert, S., & Mitchell, A. (2014). Influence of California-Style Black Ripe Olive Processing on the Formation of Acrylamide. *Journal of Agricultural and Food Chemistry*, 62(34), 8716–8721. [CrossRef]
- [32] Casado, F. J., Sanchez, A. H., & Montano, A. (2010). Reduction of acrylamide content of ripe olives by selected additives. *Food Chemistry*, 119(1), 161–166. [CrossRef]
- [33] Suman, M., Generotti, S., Cirlini, M., & Dall'Asta, C. (2019). Acrylamide reduction strategy in combination with deoxynivalenol mitigation in industrial biscuits production. *Toxins*, 11(9), 499. [CrossRef]
- [34] Graf, M., Amrein, T. M., Graf, S., Szalay, R., Escher, F., & Amadó, R. (2006). Reducing the acrylamide content of a semi-finished biscuit on industrial scale. *LWT-Food Science and Technology*, 39(7), 724–728. [CrossRef]
- [35] Yuan, Y., Chen, F., Zhao, G. H., Liu, J., Zhang, H. X., & Hu, X. S. (2007). A comparative study of acrylamide formation induced by microwave and conventional heating methods. *Journal of food science*, 72(4), C212–C216. [CrossRef]
- [36] Martins, S. I. F. S., Jongen, W. M. F., & Van Boekel, M. A. J. S. (2000). A review of Maillard reaction in food and implications to kinetic modelling. *Trends in Food Science & Technology*, 11(9–10), 364–373. [CrossRef]
- [37] Brown, R. (2003, February). Formation, occurrence and strategies to address acrylamide in food. In *FDA advisory committee meeting on acrylmaide* (Vol. 2, pp. 24–5).
- [38] Rydberg, P., Eriksson, S., Tareke, E., Karlsson, P., Ehrenberg, L., & Törnqvist, M. (2003). Investigations of factors that influence the acrylamide content of heated foodstuffs. *Journal of agricultural and food chemistry*, 51(24), 7012–7018. [CrossRef]
- [39] Generotti, S., Cirlini, M., Sarkanj, B., Sulyok, M., Berthiller, F., Dall'Asta, C., & Suman, M. (2017). Formulation and processing factors affecting trichothecene mycotoxins within industrial biscuit-making. *Food Chemistry*, 229, 597–603. [CrossRef]
- [40] Tareke, E., Rydberg, P., Karlsson, P., Eriksson, S., & Törnqvist, M. (2002). Analysis of acrylamide, a carcinogen formed in heated foodstuffs. *Journal of agricultural and food chemistry*, 50(17), 4998–5006. [CrossRef]
- [41] Becalski, A., Lau, B. P. Y., Lewis, D., & Seaman, S. W. (2003). Acrylamide in foods: occurrence, sources, and modeling. *Journal of agricultural and food chemistry*, 51(3), 802–808. [CrossRef]
- [42] Gökmen, V., Açar, Ö. Ç., Köksel, H., & Acar, J. (2007). Effects of dough formula and baking conditions on acrylamide and hydroxymethylfurfural formation in cookies. *Food chemistry*, 104(3), 1136–1142. [CrossRef]
- [43] Esposito, F., Fasano, E., De Vivo, A., Velotto, S., Sarghini, F., & Cirillo, T. (2020). Processing effects on acrylamide content in roasted coffee production. *Food chemistry*, 319, 126550. [CrossRef]
- [44] Santos, J. R., Viegas, O., Pascoa, R. N. M. J., Ferreira, I. M. P. L. V. O., Rangel, A. O. S. S., & Lopes, J. A. (2016). In-line monitoring of the coffee roasting process with near infrared spectroscopy: Measurement of sucrose and colour. *Food Chemistry*, 208, 103–110. [CrossRef]
- [45] Gökmen, V., & Palazoğlu, T. K. (2008). Acrylamide formation in foods during thermal processing with a focus on frying. *Food and bioprocess technology*, 1, 35–42. [CrossRef]
- [46] Taubert, D., Harlfinger, S., Henkes, L., Berkels, R., & Schömig, E. (2004). Influence of processing parameters on acrylamide formation during frying of potatoes. *Journal of agricultural and food chemistry*, 52(9), 2735–2739. [CrossRef]
- [47] Knol, J. J., Viklund, G. Å., Linssen, J. P., Sjöholm, I. M., Skog, K. I., & van Boekel, M. A. (2009). Kinetic modelling: A tool to predict the formation of acrylamide in potato crisps. *Food Chemistry*, 113(1), 103–109. [CrossRef]
- [48] Kita, A., Brathen, E., Knutsen, S. H., & Wicklund, T. (2004). Effective Ways of Decreasing Acrylamide Content in Potato Crisps during Processing. *Journal of Agricultural and Food Chemistry*, 52(23), 7011–7016. [CrossRef]
- [49] Bräthen, E., & Knutsen, S. H. (2005). Effect of temperature and time on the formation of acrylamide in starch-based and cereal model systems, flat breads and bread. *Food Chemistry*, 92(4), 693–700. [CrossRef]
- [50] Amrein, T. M., Bachmann, S., Noti, A., Biedermann, M., Barbosa, M. F., Biedermann-Brem, S., ... & Amadó, R. (2003). Potential of acrylamide formation, sugars, and free asparagine in potatoes: a comparison of cultivars and farming systems. *Journal of agricultural and food chemistry*, 51(18), 5556–5560. [CrossRef]
- [51] De Wilde, T., De Meulenaer, B., Mestdagh, F., Govaert, Y., Ooghe, W., Fraselle, S., ... & Verhé, R. (2006). Selection criteria for potato tubers to minimize acrylamide formation during frying. *Journal of Agricultural and Food Chemistry*, 54(6), 2199–2205. [CrossRef]
- [52] Gokmen, V., Palazolu, T. K., & Senyuva, H. Z. (2006). Relation between the acrylamide formation and time–temperature history of surface and core regions of French fries. *Journal of Food Engineering*,

- 77(4), 972–976. [CrossRef]
- [53] Deribew, H. A., & Woldegiorgis, A. Z. (2021). Acrylamide levels in coffee powder, potato chips and French fries in Addis Ababa city of Ethiopia. *Food Control*, 123, 107727. [CrossRef]
- [54] Sansano, M., De los Reyes, R., Andrés, A., & Heredia, A. (2018). Effect of microwave frying on acrylamide generation, mass transfer, color, and texture in french fries. *Food and Bioprocess Technology*, 11, 1934–1939. [CrossRef]
- [55] Oracz, J., Nebesny, E., & Zyzelewicz, D. (2011). New trends in quantification of acrylamide in food products. *Talanta*, 86, 23–34. [CrossRef]
- [56] Sansano, M., Juan-Borras, M., Escriche, I., Andres, A., & Heredia, A. (2015). Effect of Pretreatments and Air-Frying, a Novel Technology, on Acrylamide Generation in Fried Potatoes. *Journal of Food Science*, 80(5). [CrossRef]
- [57] Andrés, A., Arguelles, Á., Castelló, M. L., & Heredia, A. (2013). Mass transfer and volume changes in French fries during air frying. *Food and Bioprocess Technology*, 6, 1917–1924. [CrossRef]
- [58] Garayo, J., & Moreira, R. (2002). Vacuum frying of potato chips. *Journal of Food Engineering*, 55(2), 181–191. [CrossRef]
- [59] FAO. (2024). *Code of practice for the reduction of acrylamide in foods*. Food and Agriculture Organization of the United Nations.
- [60] U.S. Food and Drug Administration. (2023). *Reducing acrylamide in potato-based foods: Fact sheet*. Retrieved from <https://www.fda.gov/media/149436/download>
- [61] Mariotti-Celis, M. S., Cortes, P., Dueik, V., Bouchon, P., & Pedreschi, F. (2017). Application of Vacuum Frying as a Furan and Acrylamide Mitigation Technology in Potato Chips. *Food and Bioprocess Technology*, 10(11), 2092–2099. [CrossRef]
- [62] Wriston, J.C. (1985). Asparaginase. *Methods in Enzymology*, 113, 608–618.
- [63] Hendriksen, H. V., Kornbrust, B. A., Østergaard, P. R., & Stringer, M. A. (2009). Evaluating the potential for enzymatic acrylamide mitigation in a range of food products using an asparaginase from *Aspergillus oryzae*. *Journal of agricultural and food chemistry*, 57(10), 4168–4176. [CrossRef]
- [64] Ciesarová, Z., Kiss, E., & Boegl, P. (2006). Impact of L-asparaginase on acrylamide content in potato products. *Journal of Food and Nutrition Research*, 45(4), 141–146.
- [65] Mahajan, R. V., Saran, S., Kameswaran, K., Kumar, V., & Saxena, R. K. (2012). Efficient production of L-asparaginase from *Bacillus licheniformis* with low-glutaminase activity: optimization, scale up and acrylamide degradation studies. *Bioresource technology*, 125, 11–16. [CrossRef]
- [66] Pedreschi, F., Kaack, K., & Granby, K. (2008). The effect of asparaginase on acrylamide formation in French fries. *Food chemistry*, 109(2), 386–392. [CrossRef]
- [67] Zuo, S., Zhang, T., Jiang, B., & Mu, W. (2015). Reduction of acrylamide level through blanching with treatment by an extremely thermostable L-asparaginase during French fries processing. *Extremophiles*, 19, 841–851. [CrossRef]
- [68] Pedreschi, F., Mariotti, S., Granby, K., & Risum, J. (2011). Acrylamide reduction in potato chips by using commercial asparaginase in combination with conventional blanching. *LWT-food Science and Technology*, 44(6), 1473–1476. [CrossRef]
- [69] Onishi, Y., Prihanto, A. A., Yano, S., Takagi, K., Umekawa, M., & Wakayama, M. (2015). Effective treatment for suppression of acrylamide formation in fried potato chips using L-asparaginase from *Bacillus subtilis*. *3 Biotech*, 5, 783–789. [CrossRef]
- [70] Tuncel, N. B., Yilmaz, N., & Şener, E. (2010). The effect of pea (*Pisum sativum* L.)-originated asparaginase on acrylamide formation in certain bread types. *International Journal of Food Science and Technology*, 45(12), 2470–2476. [CrossRef]
- [71] Ciesarová, Z., Kukurová, K., Mikušová, L., Basil, E., Polakovičová, P., Duchoňová, L., ... & Šturdík, E. (2014). Nutritionally enhanced wheat-oat bread with reduced acrylamide level. *Quality Assurance and Safety of Crops & Foods*, 327–334. [CrossRef]
- [72] Mohan Kumar, N. S., Shimray, C. A., Indrani, D., & Manonmani, H. K. (2014). Reduction of acrylamide formation in sweet bread with L-asparaginase treatment. *Food and Bioprocess Technology*, 7(3), 741–748. [CrossRef]
- [73] Amrein, T. M., Schönbächler, B., Escher, F., & Amadò, R. (2004). Acrylamide in gingerbread: critical factors for formation and possible ways for reduction. *Journal of Agricultural and Food Chemistry*, 52(13), 4282–4288. [CrossRef]
- [74] Huang, L., Liu, Y., Sun, Y., Yan, Q., & Jiang, Z. (2014). Biochemical characterization of a novel L-Asparaginase with low glutaminase activity from *Rhizomucor miehei* and its application in food safety and leukemia treatment. *Applied and environmental microbiology*, 80(5), 1561–1569. [CrossRef]
- [75] Anese, M., Quarta, B., & Frias, J. (2011). Modelling the effect of asparaginase in reducing acrylamide formation in biscuits. *Food chemistry*, 126(2), 435–440. [CrossRef]
- [76] Vass, M., M Amrein, T., Schonbachler, B., Escher, F., & Amado, R. (2004). Ways to reduce the acrylamide formation in cracker products. *Czech Journal of Food Sciences*, 22(SI-Chem. Reactions in Foods V), S19–S21. [CrossRef]
- [77] Muttucumaru, N., Powers, S. J., Elmore, J. S., Dodson, A., Briddon, A., Mottram, D. S., & Halford, N. G. (2017). Acrylamide-forming potential of potatoes grown at different locations, and the ratio of free asparagine to reducing sugars at which free

- asparagine becomes a limiting factor for acrylamide formation. *Food chemistry*, 220, 76-86. [CrossRef]
- [78] Liyanage, D. W., Yevtushenko, D. P., Konschuh, M., Bizimungu, B., & Lu, Z. X. (2021). Processing strategies to decrease acrylamide formation, reducing sugars and free asparagine content in potato chips from three commercial cultivars. *Food control*, 119, 107452. [CrossRef]
- [79] Elmore, J. S., Koutsidis, G., Dodson, A. T., Mottram, D. S., & Wedzicha, B. L. (2005). Measurement of Acrylamide and Its Precursors in Potato, Wheat, and Rye Model Systems. *Journal of Agricultural and Food Chemistry*, 53(4), 1286–1293. [CrossRef]
- [80] Crawford, L. M., Kahlon, T. S., Chiu, M. C. M., Wang, S. C., & Friedman, M. (2019). Acrylamide content of experimental and commercial flatbreads. *Journal of food science*, 84(3), 659-666. [CrossRef]
- [81] Banchemo, M., Pellegrino, G., & Manna, L. (2013). Supercritical fluid extraction as a potential mitigation strategy for the reduction of acrylamide level in coffee. *Journal of Food Engineering*, 115(3), 292-297. [CrossRef]
- [82] Tuta, S., Palazoglu, T. K., & Gokmen, V. (2010). Effect of microwave pre-thawing of frozen potato strips on acrylamide level and quality of French fries. *Journal of food engineering*, 97(2), 261-266. [CrossRef]
- [83] Liu, H., Li, X., & Yuan, Y. (2020). Mitigation effect of sodium alginate on acrylamide formation in fried potato chips system based on response surface methodology. *Journal of Food Science*, 85(8), 2615–2621. [CrossRef]
- [84] Garmakhany, A. D., Mirzaei, H. O., Nejad, M. K., & Maghsudlo, Y. (2008). Study of oil uptake and some quality attributes of potato chips affected by hydrocolloids. *European journal of lipid science and technology*, 110(11), 1045-1049. [CrossRef]
- [85] Hua, X., Wang, K., Yang, R., Kang, J., & Yang, H. (2015). Edible coatings from sunflower head pectin to reduce lipid uptake in fried potato chips. *LWT-Food Science and Technology*, 62(2), 1220-1225. [CrossRef]
- [86] Champrasert, O., Chu, J., Meng, Q., Viney, S., Holmes, M., Suwannaporn, P., & Orfila, C. (2021). Inhibitory effect of polysaccharides on acrylamide formation in chemical and food model systems. *Food Chemistry*, 363, 130213. [CrossRef]
- [87] Bartkiene, E., Bartkevics, V., Pugajeva, I., Krungleviciute, V., Mayrhofer, S., & Domig, K. (2017). The contribution of P. acidilactici, L. plantarum, and L. curvatus starters and L-(+)-lactic acid to the acrylamide content and quality parameters of mixed rye-Wheat bread. *Lwt*, 80, 43-50. [CrossRef]
- [88] Dastmalchi, F., Razavi, S. H., Faraji, M., & Labbafi, M. (2016). Effect of Lactobacillus casei- casei and Lactobacillus reuteri on acrylamide formation in flat bread and Bread roll. *Journal of food science and technology*, 53(3), 1531–1539. [CrossRef]
- [89] Nguyen, H. T., Van Der Fels-Klerx, H. J. (Ine)., & Van Boekel, M. A. J. S. (2017). Acrylamide and 5-hydroxymethylfurfural formation during biscuit baking. Part II: Effect of the ratio of reducing sugars and asparagine. *Food Chemistry*, 230, 14–23. [CrossRef]

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